

Hepatoprotective Effect of *Stevia rebaudiana* Bertoni Leaf Extract in CCl₄-Induced Liver Injury in Albino Rats

Karthik Mohan^{1*} • Jeyachandran Robert²

¹ Department of Biochemistry, St. Joseph's College (Autonomous), Tiruchirapalli -620002, Tamil Nadu, India

² Department of Plant Biology and Plant Biotechnology, Tiruchirapalli -620002, Tamil Nadu, India

Corresponding author: * km80_profsjc@yahoo.co.in

ABSTRACT

The efficacy of aqueous leaf extract of *Stevia rebaudiana*, a non-calorific natural sweetener was evaluated against carbon tetrachloride (CCl₄)-induced liver damage in albino rats. Hepatotoxicity was induced by *i.p.* administration of 30% CCl₄ suspended in olive oil (1 ml/kg of body weight). After the treatment regimen animals were sacrificed and liver enzymes AST, ALT, ALP, and LDH were assayed in serum along with total protein and antioxidant status was assessed in liver tissues by determining the activities of SOD, CAT, GPx and GSH. The lipid peroxidation levels were also assessed. The observations suggested that pre-treatment of the natural sweetener could possibly protect CCl₄-induced liver damage and that the exerted effect is connected with its antioxidant and hepatoprotective nature.

Keywords: antioxidant, liver injury, natural sweetener, stevioside

INTRODUCTION

Stevia rebaudiana Bertoni, also termed 'sweet leaf' or *seeni tulusi* (in Tamil), belongs to the *Asteracea* family, native to subtropical and tropical South America (Braguini *et al.* 2003; reviewed in Meireles *et al.* 2006). The leaf extract of *S. rebaudiana* is known for its sweetness which is 300 times sweeter than table sugar and has been used as a sweetening agent for many centuries. Even today in the eastern countries *Stevia* is widely used as a food additive. The glycosides of *Stevia* were identified as the sweetening component of these leaves and stevioside is the major component (Bondarev *et al.* 2002). Clinical studies conducted earlier revealed the hypoglycemic and antihypertensive effect of this plant (Chan *et al.* 2000; Hsieh *et al.* 2003; Gregersen *et al.* 2004). These findings encouraged the use of *Stevia* by diabetic patients who crave for sweet taste and suffer from the side effects of artificial sweetening agents such as aspartame, saccharin and cyclamate (Usami *et al.* 1980). Few artificial sweeteners are carcinogenic and severally affect the normal liver functioning, although researchers raised concern over the use of *Stevia* since they believed that it might be toxic to the liver (Kelmer *et al.* 1985; Braguini *et al.* 2003).

There is thus a need to test the effect of *Stevia* on the liver. In this study, the efficacy of the aqueous leaf extract of *S. rebaudiana* was tested in CCl₄-induced liver damage in albino rats and if found to possess a hepatoprotective effect then *Stevia* would remain important for diabetics without the fear of chronic toxicity of artificial sweeteners.

MATERIALS AND METHODS

Plant material

S. rebaudiana leaves were purchased from commercial sellers, and were authenticated by The Raphinat Herbarium, St. Joseph's College, Trichy. They were shade dried at 37°C for 7 days. The dried material was powdered and stored in a dry container at room temperature.

Preparation of extract

100 g of dry leaf powder was suspended in water at a ratio of 1:600 (w/v) and stirred at 2000 rpm with a magnetic stirrer overnight at 37°C. The process was repeated in triplicate and then the extract, which was filtered using a fine muslin cloth, was stored in the refrigerator for further use.

The analytical grade chemicals used were purchased from Himedia Chemicals (India).

Animals

Male albino rats weighing about 100-150 g were obtained from the Tamil Nadu Veterinary and Animal Sciences University, Madhavaram, Chennai. The animals were housed in a well ventilated animal room of the Department of Biochemistry, St. Joseph's College, Trichy, under a 12-h light/dark cycle. The animals were fed with standard pellet diet and water *ad libitum*. The use of animals was in accordance with the rules of the Institutional animal ethical committee.

Experimental design

The animals were divided into four groups of 6 animals each and treated as follows: Group I animals served as normal control, Group II animals constituted the hepatotoxic group, which received *i.p.* administration of 30% CCl₄ suspended in olive oil (Rocchi Olive oil, Hemraj, India) (1 ml/kg of body weight) (Castro *et al.* 1974), Group III and IV animals were treated with the *Stevia* leaf extract (SLE) for 10 days and for Group IV animals CCl₄ was given as in group II on the 11th day after pretreatment with the extracts. The animals were sacrificed on the 12th day by cervical decapitation. Blood was collected and serum was separated and stored at 4°C for further analysis. Serum marker enzymes analyzed were aspartate transaminase (AST) (King 1965), alanine transaminase (ALT) (King 1965), alkaline phosphatase (ALP) (Kind and King 1954), lactate dehydrogenase (LDH) (Ulmer *et al.* 1956), total serum protein (TSP) (Lowry *et al.* 1951). Antioxidant status was analyzed from the levels of glutathione (GSH) (Beutler and Kelley 1963), lipid peroxidation (LP) (Stocks and Dormandy 1971) and activities of glutathione peroxidase (GPx) (Rotruck *et al.*

1973), catalase (CAT) (Bergmeyer *et al.* 1974) and superoxide dismutase (SOD) (Roos *et al.* 1979) of the liver tissue. The results obtained were statistically analyzed by the student's *t*-test for significance ($P < 0.05$; < 0.001) between groups (Bennett and Franklin 1967).

RESULTS AND DISCUSSION

The levels of the marker enzymes ALP, AST, ALT, LDH and TSP of all the four groups experimental and control groups are shown in **Table 1**. The levels of these parameters, except for serum proteins, showed a significant increase in their levels in animals belonging to group II, the toxicity-induced group whereas the protein values decreased in the CCl₄-induced animals. The level was found to be near normal with no significant difference when compared with the control group animals and the treatment group (IV) showed marked elevation in the total protein levels and decrease in enzyme levels. The results of the antioxidant enzymes (**Table 2**) show that administration of CCl₄ decreased the antioxidant enzyme levels and a significant increase was observed in the treatment group. The lipid peroxidation assayed by the TBARS method also showed similar results and animals treated with the plant extract alone maintained normalcy to a greater extent.

The present study was conducted to determine the anti-hepatotoxic effect of *S. rebaudiana* leaf extract in albino rats. CCl₄ was used to induce hepatic damage in rats since it is known to produce acute hepatocellular injury with centrilobular necrosis and steatosis (Recknagel *et al.* 1989). It is a highly toxic chemical and its adverse effects on liver have (Recknagel 1967) been known for years (and hence it used as an ideal model to induce hepatic cirrhosis). CCl₄ metabolites such as trichloromethyl peroxyradical are also toxic to the liver (Mehandale *et al.* 1986). On induction of CCl₄ the levels of marker enzymes increased significantly compared to the normal control group I (**Table 1**). Enzyme activities decreased considerably in treatment group IV compared to the induced group II. The activities of these enzymes remained near normal and increased slightly in a few cases in Group III (**Table 1**). The increased levels of AST, ALT, ALP, and LDH in serum indicate damage to hepatic cells (Wolf 1999). Fluctuations in AST levels indicate a loss of cell integrity, while variations in the activity of ALT and LDH indicate increased permeability and cellular leakage (Sreepriya *et al.* 2001). Similarly, the activities of AST, ALT,

ALP and LDH in group IV animals were found to decrease significantly compared to the induced group II when treated for experimentally induced myocardial infarction (Mohan and Robert 2007). This shows that pretreatment of *S. rebaudiana* leaf extract is capable of preventing cellular damage induced by CCl₄. The decrease in the total protein level in the induced group might be because of the decrease in the liver's ability to synthesize proteins (Ahmed *et al.* 2000). But the animals belonging to Group IV exerted a marked increase in total protein levels in serum, which is an indication of the regeneration of the damaged liver tissues and that the *S. rebaudiana* leaf extract was capable of rendering some resistance to hepatic tissues to withstand the degenerative nature of CCl₄. The antioxidant status of the *S. rebaudiana* leaf extract was evident from an increase in the levels of antioxidant enzymes (Mohan and Robert 2009). In the CCl₄-induced groups the levels of the antioxidant enzymes SOD, CAT, GPx, GSH were found to decrease and lipid peroxidation levels increased in Group II (**Table 2**). In CCl₄-induced groups lipid peroxidation was evident and might be because of changes in the cell membrane (Ohta *et al.* 1995). The decrease in the levels in group IV indicated the anti-lipid peroxidative effect of *S. rebaudiana*. SOD acts as a cellular defense element against potentially harmful effects of superoxide radicals by catalyzing the dismutation of these ions (Mohanan *et al.* 1997) (**Table 2**). GST increases the conjugation of free radicals and lipid hydroperoxides to GSH and enhances its excreatability (Beckett and Hayes 1987). Catalase was also involved in ROS scavenging. The decline in the activities of SOD, CAT, GPx and GSH in the induced group and significant increase in the treated group indicates the oxidative damage produced by CCl₄ to liver (Paduraru *et al.* 1996) and the protective effect rendered by *S. rebaudiana* leaf extract to the treated animals (**Table 2**). These protective effects exerted by *S. rebaudiana* might be because of its glycosides.

CONCLUSION

The present study demonstrated the hepatoprotective and antioxidant effect of *S. rebaudiana* leaf extract in experimentally induced hepatic injury in albino rats. Further research is to be carried out to determine the active principle and dose dependent effect.

Table 1 Effect of *Stevia rebaudiana* leaf extract on liver marker enzymes in the serum of control and experimental animals.

Parameters	Group I	Group II	Group III	Group IV
ALT (U/ml)	45.01 ± 1.9	115.67 ± 8.93 ⁺	47.24 ± 1.66	80.13 ± 5.10 ^{**}
AST (U/ml)	53.05 ± 3.16	165.26 ± 10.2 ⁺	54.98 ± 2.10	125.30 ± 10.42 ^{**}
ALP (KA units)	48.32 ± 2.11	99.65 ± 5.42 ⁺	50.65 ± 3.12	63.75 ± 4.27 [*]
LDH (IU/dl)	100.01 ± 1.02	151.68 ± 10.97 ⁺	103.88 ± 3.32	140.64 ± 9.98 [*]
TSP (mg/ml)	52.45 ± 1.48	48.12 ± 3.46 ⁺	53.59 ± 2.15	50.55 ± 1.50 ^{**}

Group I = Normal control, Group II = CCl₄ toxicity induced, Group III = *Stevia* leaf extract alone treated, Group IV = CCl₄ induced group pretreated with *Stevia* leaf extract.

Group III were compared with Groups I and IV were compared with Group II.

Values are mean ± SEM for 6 animals in each group

* $P < 0.05$; ** $P < 0.001$; as compared with CCl₄-induced groups.

⁺ $P < 0.001$ as compared with normal group.

ALT, alanine transaminase; ALP, alkaline phosphatase; AST, aspartate transaminase; LDH, lactate dehydrogenase; TSP, total serum protein

Table 2 Effect of *Stevia rebaudiana* leaf extract on antioxidant status in the liver of control and experimental animals.

Parameters	Group I	Group II	Group III	Group IV
GSH (µg/mg of protein)	15.02 ± 1.3	10.59 ± 1.65 ⁺	15.43 ± 0.2	13.72 ± 1.0 [*]
LP (nmoles of malondialdehyde (MDA) formed/mg of protein / hr)	120.20 ± 20.1	186.32 ± 12.7 ⁺	121.06 ± 0.97	140.33 ± 7.6 ^{**}
GPx (µg of glutathione consumed/min/mg protein)	17.00 ± 1.0	15.12 ± 1.02 ⁺	16.92 ± 0.05	16.02 ± 0.05 [*]
CAT (unit/min/mg of protein)	34.39 ± 1.50	15.02 ± 0.38 ⁺	34.06 ± 0.04	21.15 ± 0.81 ^{**}
SOD (Unit/min/mg of protein)	2.45 ± 0.05	1.80 ± 0.04 ⁺	2.31 ± 0.06	1.82 ± 0.09

Group I = Normal control, Group II = CCl₄ toxicity induced, Group III = *Stevia* leaf extract alone treated, Group IV = CCl₄ induced group pretreated with *Stevia* leaf extract.

Group III were compared with Groups I and IV were compared with Group II.

Values are mean ± SEM for 6 animals in each group.

* $P < 0.05$; ** $P < 0.001$; as compared with CCl₄-induced groups.

⁺ $P < 0.001$ as compared with normal group.

CAT, catalase; GPx, glutathione peroxidase; GSH, glutathione; LP, lipid peroxidation; SOD, superoxide dismutase

REFERENCES

- Ahmed S, Rahman A, Alam A, Saleem M, Athar M, Sultana S (2000) Evaluation of the efficacy of *Lawsonia alba* in the alleviation carbon tetrachloride induced oxidative stress. *Journal of Ethnopharmacology* **69**, 157-164
- Beckett GJ, Hayes JD (1987) Glutathione S-transferase measurements and liver disease in man. *Journal of Clinical Biochemistry and Nutrition* **2**, 1-24
- Bennett CA, Franklin NL (1967) *Statistical Analysis in Chemistry and Chemical Industry*, John Wiley and Sons, Inc, New York, pp 133-134
- Bergmeyer H, Gawehn K, Grasse M (1974) *Enzymes as Biochemical Reagents* In: Bergmeyer HV (Ed) *Methods of Enzymatic Analysis*, Academic Press, New York, pp 438-448
- Beutler E, Kelley BM (1963) The effects of sodium nitrate on red cell glutathione. *Experientia* **19**, 96-97
- Bondarev N, Reshetnyak O, Nosov A (2002) Features of development of *Stevia rebaudiana* shoots cultivated in the roller bioreactor and their production of steviol glycosides. *Planta Medica* **68**, 759-762
- Braguini WL, Gomes MA, Oliveira BH, Carnieri EG, Rocha ME, Oliveira MB (2003) Activity of isosteviol lactone on mitochondrial metabolism. *Toxicology Letters* **143**, 83-92.
- De Castro JA, De Ferreyra EC, Fenoos OM, Gillette JR (1974) Prevention of carbon tetrachloride induced necrosis by inhibitors of drug metabolism. *Biochemistry and Pharmacology* **23**, 295-302
- Chan P, Tomlinson B, Chen YJ, Liu JC, Hsieh MH, Cheng JT (2000) Double-blind placebo- controlled study of the effectiveness and tolerability of oral stevioside in human hypertension. *British Journal of Clinical Pharmacology* **50**, 215-220
- Gregersen S, Jeppesen PB, Holst JJ, Hermansen K (2004) Antihyperglycemic effects of stevioside in type 2 diabetes. *Metabolism* **53**, 73-76
- Hsieh MH, Chan P, Sue YM, Liu JC, Liang TH, Huang TY, Tomlinson B, Chow MS, Kao PF, Chen YJ (2003) Efficacy and tolerability of oral stevioside in patients with mild essential hypertension: a two-year, randomized, placebo-controlled study. *Clinical Therapeutics* **25**, 2797-2808
- Kelmer Bracht A, Alvarez M, Bracht A (1985) Effect of *Stevia rebaudiana* natural products on rat liver mitochondria. *Biochemistry and Pharmacology* **34**, 873-882
- Kind PRM, King EJ (1954) Estimation of plasma phosphatase by determination of hydrolysed phenol with amino antipyrine. *Journal of Clinical Pathology* **7**, 322-326
- King J (1965) *Practical Clinical Enzymology: The Hydrolases - Acid and Alkaline Phosphates*, D Van Nostrand Company, London, pp 191-208, 363-367
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951) Protein measurement with the folin phenol reagent. *The Journal of Biological Chemistry* **193**, 265-275
- Mehandale HM, Roth RA, Gandolfi AJ, Klaunig JE, Le-masters JJ, Curtis LR (1986) Novel mechanisms in chemically induced hepatotoxicity. *Federation of American Societies for Experimental Biology Journal* **104**, 826-829
- Meireles MAA, Wang G-M, Hao Z-B, Shima K, Teixeira da Silva JA (2006) *Stevia (Stevia rebaudiana Bertoni): futuristic view of the sweeter side of life*. In: Teixeira da Silva JA (Ed) *Floriculture, Ornamental and Plant Biotechnology: Advances and Topical Issues* (Vol IV), Global Science Books Ltd., Isleworth, UK, pp 416-425
- Mohan K, Robert J (2007) Effect of *Stevia rebaudiana* leaf extract in experimentally induced Myocardial infarction in male albino rats. *Journal of Tropical Medicinal Plants* **7** (1), 63-67
- Mohan K, Robert J (2009) Protective effect of *Stevia rebaudiana* Bertoni on cadmium-induced nephrotoxicity in male albino rats. *International Journal of Biomedical and Pharmaceutical Sciences* **3**, in press
- Mohan PV, Rathinam K, Devi KS (1997) Effect of *Solanum trilobatum* on ultra violet induced damage and super oxide production. *Indian Journal of Pharmacology* **29**, 129-131
- Ohta Y, Sasaki E, Nishida K, Kobayashi T, Nagata M, Ishigur I (1995) Preventive effect of dia-saiko-to (da-chai-hutang) extract on disrupted hepatic active oxygen metabolism in rats with carbon tetrachloride induced liver injury. *American Journal of Clinical Medicine* **23**, 53-64
- Paduraru I, Saramet A, Danila GH, Nichifor M, Jerca L, Iacobovici A (1996) Antioxidant action of a new flavonic derivative in acute carbon tetrachloride intoxication. *European Journal of Drug Metabolism and Pharmacokinetics* **21**, 1-6
- Recknagel RO (1967) Carbon tetrachloride hepatotoxicity. *Pharmacology Review* **19**, 145-195
- Recknagel RO, Glende EA Jr., Dolak JA, Walker RL (1989) Mechanisms of carbon tetrachloride toxicity. *Pharmacology and Therapeutics* **43**, 139-154
- Roos D, Weening RS, Voetman AA, Van Schaik ML, Bot AA, Meerhof LJ (1979) Protection of phagocytic leucocytes by endogenous glutathione: studies in a family with glutathione reductase deficiency. *Blood* **53**, 851-866
- Rotruck, JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Haekstra WG (1973) Selenium: biochemical role as component of glutathione peroxidase. *Science* **179**, 588-590
- Sreepriya M, Devaki T, Balakrishnana K, Apparannantham T (2001) Effect of *Indigofera tinctoria* Linn. on liver antioxidant defense system during D-galactosamine/endotoxin-induced acute hepatitis in rodents. *Indian Journal of Experimental Biology* **39**, 181-184
- Stocks J, Dormandy TL (1971) Autoxidation of human red cell lipids induced by hydrogen peroxide. *British Journal of Haematology* **20**, 95-111
- Ulmer DD, Valle BL, Wacker WE (1956) Metalloenzymes and myocardial infarction. Malic and lactic dehydrogenase activities and zinc concentration in serum. *New England Journal of Medicine* **225**, 449-456
- Usami M, Seino Y, Takai J (1980) Effect of cyclamate sodium, saccharin sodium and stevioside on arginine-induced insulin and glucagons secretion in the isolated perfused rat pancreas. *Hormone and Metabolic Research* **12**, 705-706
- Wolf PL (1999) Biochemical diagnosis of liver disease. *Indian Journal of Clinical Biochemistry* **14**, 59-65